

CAPE ELEUTHERA INSTITUTE SUSTAINABLE DEVELOPMENT

INTRODUCTION

Global climate change is an issue that impacts a wide range of marine ecosystems, thereby generating unnatural and potentially harmful behavioral and physiological responses from the organisms of these ecosystems (Karl and Trenberth 2003). Climate change is largely influenced by human interference with the earth's atmospheric composition (Karl and Trenberth 2003). The effects of a global warming trend are evident due to recent changes in global weather patterns, temperatures, and precipitation levels (Brierley and Kingsford 2009). Projected warming patterns for the future raise even more concern about the ecological and also socio-economic consequences of climate change (Walther et al. 2002).

One of the major ecological groups impacted by climate change is the marine environment. Little research has been conducted on tropical marine ecosystems, and they are therefore the hardest aquatic habitats to evaluate in terms of tropical fishes' reactions to climate change (Roessig et al. 2004). One study highlighted the effects of global warming on fish physiology. This experiment on tropical coral reef fish in Australia found that combined variables of augmented water temperature and acidity increased the oxygen consumption and mortality rates of fish. When dealing with stressors, fish allocate less energy to basic survival behaviors such as foraging, escaping predators, and reproducing (Munday et al. 2009).

Bonefish are critical contributors to the tropical flats ecosystems of the Bahamian Archipelago (Danylchuk et al.). In addition, recreational bonefish angling generates 141 million dollars annually for the Bahamian economy (Danylchuk et al.). Despite the ecological and economic importance of bonefish, relatively little information is known about this species. The purpose of this study was to examine how climate change affects the metabolic rates of bonefish (Albula spp.). The hypothesis was that if exposed to increased temperature and acidity treatments, bonefish would have escalated oxygen consumption rates and stress levels.



Figure 1. Seine net encircled around a school of bonefish for collection and return to CEI wet labs.



Figure 2. A bonefish angler in a Bahamian flats environment.

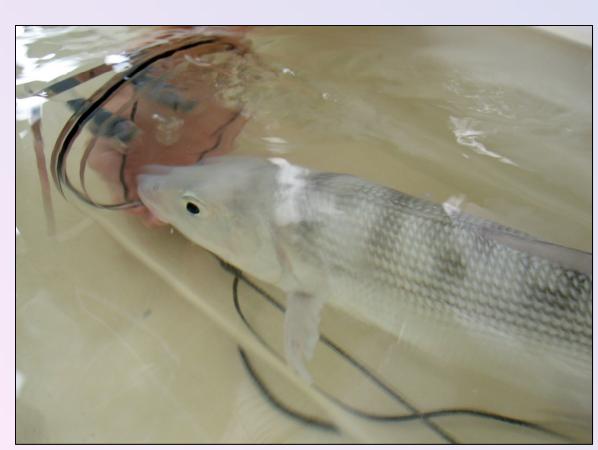
METHODOLOGY

This study was conducted at the Cape Eleuthera Institute (CEI) on Eleuthera, The Bahamas during the fall of 2010. Using a seine net, bonefish (Albula spp.) from local tidal creeks were caught and transported to the CEI wet labs. The water in the acclimation tank was adjusted over a 24 hour period from normal ocean temperature and acidity levels (30°C and 8.1pH, respectively) to higher levels of 33°, 34° or 7.8, 7.6, 7.4 pH. Heaters were immersed in the water to increase temperature, and carbon dioxide was bubbled into the water to increase acidity. Conditions were regulated by means of a pH controller monitor and a thermostat

After an acclimation period of 7 days, the weights and volumes of fish were taken, as these two factors affect the metabolic rate of each fish. The bonefish were then placed in respirometry chambers and their rates of oxygen consumption were measured (Figure 3). The respirometer consists of four individual chambers placed in a tank, and each chamber is outfitted with an oxygen probe that allows for simultaneous monitoring of oxygen consumption from 4 fish (Figure 4). Data on the standard metabolic rate of bonefish was collected overnight when there were few disturbances (Munday et al. 2009).

Immediately following the respirometry period, the bonefish were moved to an exercise tank where predatory activity was simulated (Figure 5). The amount of time that it took each fish to become exhausted was measured. Bonefish were then tagged and released.

One-way ANOVA tests were performed in order to determine whether the oxygen consumption and swimming performance of control bonefish differed significantly from that of the treatment bonefish. A one-way ANOVA was also used to determine if there were significant differences between fish lengths between different treatment groups.



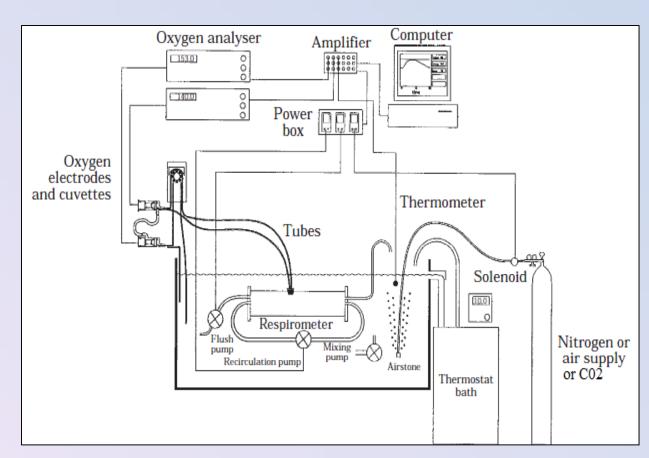
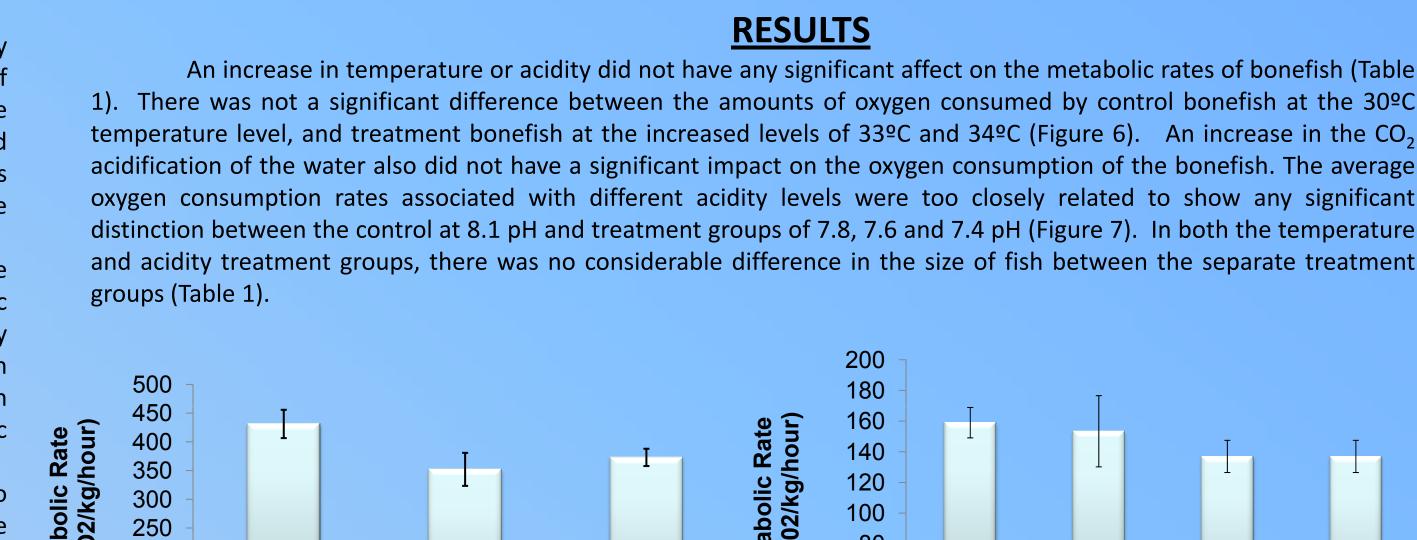


Figure 3. Bonefish in a respirometry chamber.

Figure 4. Diagram representing the setup of the respirometry equipment.

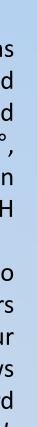
The metabolic response of bonefish (Albula spp.) in the Bahamian Archipelago to increased ocean temperature and acidity caused by global climate change

Student researchers: Ellen Doughty, CJ Easton, Chris Daniell, Noah Boskey, Hannah Leeman, and Heather Seeley Project advisors: Aaron Shultz and Liane Nowell



5

34°C





50 33°C Contro Treatment Figure 6. The average oxygen consumption rates of each temperature treatment group of bonefish. The error bars represent ± 1 SE. Sample sizes are shown within graph bars.

3

200

150

100

Met (mg

Figure 7. The average oxygen consumption rates of each acidity treatment group of bonefish. The error bars represent \pm 1 SE. Sample sizes are shown within graph bars.

7.8pH

The data collected during the exercise period for the bonefish revealed similar results to the findings on the fishes' oxygen consumption. There was no significant variation between the number of seconds that it took the control bonefish and the treatment fish of both temperature and acidity treatments to become exhausted while swimming, as supported by statistical analyses (Table 1, Figures 8 and 9, respectively).

20

180

160

Contro

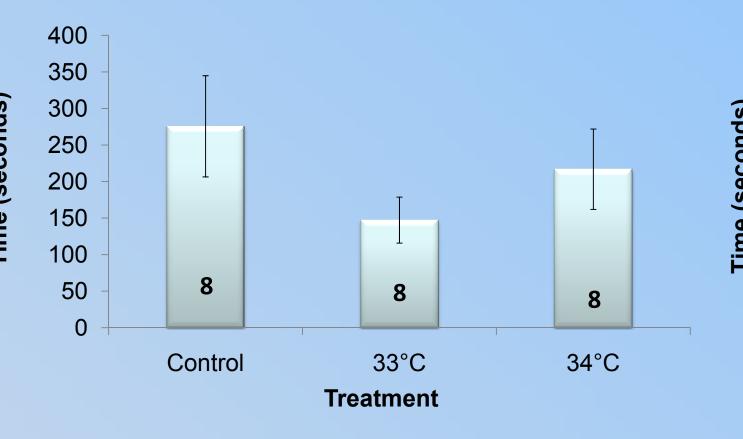


Figure 8. The average time (seconds) that it took each temperature treatment group of bonefish to become exhausted during the exercise period. The error bars represent \pm 1 SE. Sample sizes are shown within graph bars.

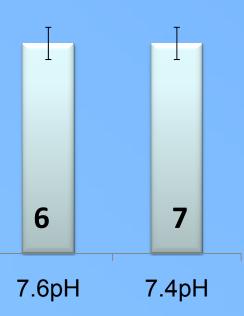
Figure 9. The average time (seconds) that it took each acidity treatment group of bonefish to become exhausted during the exercise period. The error bars represent \pm 1 SE. Sample sizes are shown within graph bars.

Table 1. Results from statistical tests showing differences in dissolved O_2 concentrations between treatment groups

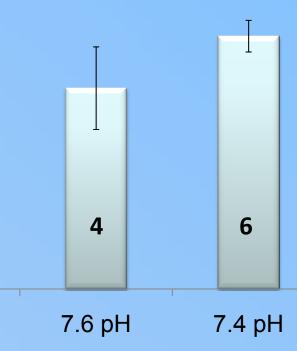
Treatment	F value	P value
Temperature		
(respirometry data)	F _{2,8} = 0.2974	P > 0.7506
рН		
(respirometry data)	F _{3,22} = 0.7678	P > 0.5243
Temperature		
(swimming performance)	F _{2,21} = 1.7359	P > 0.05
рН		
(swimming performance)	F _{3,21} = 0.5313	P > 0.05



Figure 5. Bonefish during the predation simulation.



Treatmen



Treatment

DISCUSSION

According to the results of this study, increased water temperatures and acidities produced no significant change in the oxygen consumption rates of bonefish. The exercise data showed that the swimming performance of the bonefish was also not impacted by exposure to the treatment variables.

Treatment bonefish were expected to exhibit increased oxygen consumption rates when exposed to temperatures and acidities higher than ambient ocean levels. The results of the study do not support this hypothesis. These results also disagree with findings of related studies, such as the experiment conducted on coral reef fish in Australia (Munday et al. 2009).

The conclusions of this study suggest that bonefish are very adaptable to variations in natural water conditions caused by anthropogenic climate change. This high tolerance of bonefish can be attributed to the relatively "harsh" environments that these fish inhabit. The shallow water and near-shore location of mangrove flats ecosystems expose these habitats to perturbations like acid rain and changes in temperature due to tidal shifts. Consequently, the marine species of flats ecosystems, namely bonefish, have developed a high resilience to the disturbances of climate change (Alongi 2007).

Further research needs to be conducted on tropical fishes' oxygen consumption in order to improve understanding of the impacts of climate change on these fish. Future studies could include focusing on a wider variety of tropical fish species and locations, increasing the magnitude of treatment variables, or exposing fish to combined treatment variables. Future experiments should also increase the number of fish in each treatment group, as the bonefish sample sizes for this study were much smaller than ideal.



Figure 10. A bonefish in the shallow flats environment.



Figure 11. Mangrove flats ecosystem in the Bahamian Archipelago.



Figure 12. Snorkeling in a local flats tidal creek.

ACKNOWLEDGEMENTS

We would like to extend our thanks to the following individuals and institutions for all of their support and insight that went into this research project:

> David Philipp and Cory Suski The Cape Eleuthera Institute The Island School The University of Illinois

LITERATURE CITED Alongi, D.M. 2007. Mangrove forests: Resilience, protection from tsunamis, and responses to global climate change. Estuarine, Coastal, and Shelf Science 76: 1-13. Brierley, A.S. and Kingsford, M.J. 2009. Impacts of climate change on marine organisms and ecosystems. Current Biology 19: R602-R614. Danylchuk, A.J., Danylchuk, S.E., Cooke, S.J., Goldberg, T.L., J. Koppelman, and Philipp, D.P. Ecology and management of bonefish (Albula Spp) in the Bahamian Archipelago. Ecology and Management of Bahamian Bonefish 1-25. Karl, T.R. and Trenberth, K.E. 2003. Modern Global Climate Change. Science 302: 1719-1723. Munday, P.L., Crawley, N.E., and Nilsson, G.E. 2009. Interacting effects of elevated temperature and ocean acidification on the aerobic performance of coral reef fishes. Marine Ecology Progress Series 388: 235-242. Roessig, J.M., Woodley, C.M., Cech, J.J. Jr., and Hansen, L.J. 2004. Effects of global climate change on marine and estuarine fishes and fisheries. Reviews in Fish Biology and Fisheries 14: 251–275. Schurmann, H. and Steffensen, J.F. 1997. Effects of temperature, hypoxia and activity on the metabolism of juvenile Atlantic cod. Journal of Fish Biology 50: 1166-1180. Walther, G., Post, E., Convey, P., Menzel, A., Parmesank, C., Beebee, T.J.C., Fromentin, J., Hoegh-Guldberg, O., and Bairlein, F. 2002. Ecological Responses to Recent Climate Change. Nature 416: 389-395.